# Isolation and Identification of *Clostridium difficile* from Antibiotic- Associated Diarrhea and Colitis in Iraqi children, using API20A Anaerobic system.

Mona Turkey AL-Mossawei

Luma Yousif Mahdi

Biology Department, College of Science for Women Baghdad University.

## Abstract

Clostridium difficile were isolated by 51 (21.25%) from a total of 240 stool sample collected from children having diarrhea and colitis at ages from after birth to 15 years old from Baghdad hospitals, who were under treatment with broad spectrum antibiotics, and Sample were taken during the period of first of June 2013 till the end of april 2014. Also 80 stool samples were collected from healthy ,non diarrhea children as controls. Morphological and API 20A tests were used for the identification of this bacterium to evaluate the performance of the API 20A anaerobe system in definitively identifying strains of C. difficile. This study shows that females were more infected than males .Overall positivity was (21.25%) in this study group compared to controls (P<0.05). The isolation percent from age <5 years was (18.75%), while it in age>5 years was(2.5%). It was found that this percent decreased with age get older.

Key words: API20A KIT, *Clostridium difficile*, Antibiotic-Associated Diarrhea(AAD), Colitis. **Introduction** 

*Clostridium difficile* is the most common cause of Antibiotic-Associated Diarrhea and Colitis, usually occurring after exposure to antibiotics[1]. During past few years,

*C. difficile* infection has become more frequent, more severe, more refractory to standard treatment, and more likely to relapse [2]. *C. difficile* remains the most important cause of healthcare-associated diarrhea and is increasingly important as a community pathogen. The strains of *C. difficile* 

مجلة كلية التربية الأساسية - 57 - المجلد 21- العدد 87- 2015

with hyper virulent have been reported for after the use of nearly all systemic antibacterial agents worldwide[3].

The decreased effectiveness of metronidazole relative to vancomycin in the treatment of *C.difficile* has been demonstrated[4]. The pathogenic effects of C. difficile are mucosal damage to the colon that is caused by the production of toxin A (308 kDa) and/or toxin B (270 kDa) [5]. Many infants are colonized by toxigenic or non -toxigenic strains during the first two years of their life. A study was sparked by the finding of stool positive for C. difficile whose mortality was linked to sudden infant death syndrome, showed significantly greater colonization in newborns fed on formula 71% than in breast-fed infants 7%. [6, 7]. C. difficile is a spore-forming, obligate anaerobic, Gram positive bacillus and is usually acquired either from the environment or via the fecal- oral [ 8]. C. difficile is a bacterium of ubiquitous nature whose spore route ability renders it highly resistant to environmental and food producing associated stresses[9]. CDI has become an increasingly production common infection and has shown an increase in severity over the past Since not all strains of C. difficile form toxins and several years[10]. approx. 2 % healthy adults and up to 50% of children under the age of 2 with C. difficile. [11]. Previous study in Iraq years may be parasitized isolated 14%, diarrhea8%, and healthy 6% [12], and due to the importance of detection of *C.difficile* infection especially in children under 5 years, and because of rare of previous study in Iraq of this bacteria , suggested to do this study.

# Material & Methods

## Sample:

335 stool sample were collected from children suffering from Antibiotic-Associated Diarrhea( AAD) and Colitis, after exposure to antibiotics ,cases at ages from after birth to 15 years old in Baghdad hospitals, and 240 samples of them was chosen from non-fungus and nonparasite fesses . Sample were taken during the period of first of June 2013 till the end of April 2014. Also80 stool samples were collected from healthy ,non diarrhea children as control .

#### Media:

*C. difficile* agar base : for the isolation of *C. difficile* when used with supplements .(SIGMA-ALDRICH,USA) .CCFA (Cycloseren-Cefoxitin-Fructose Agar.(As a selective media).And 7%horse blood[13].

Technique for Alcohol Shock Treatment[13, 14].:

مجلة كلية التربية الأساسية \_ 58 \_ المجلد 21- العدد 87- 2015

Mix equal parts of industrial absolute alcohol and the fecal specimen. Homogenize using a vortex mixer. Leave at room temperature for 1 hr. Inoculate on to *C. Difficile* Selective Agar and Incubate at 37°C in a conventional anaerobic gas jar. The use of the Oxoid Anaerobic Jar with Anaerogen (AN0025,OXOID,UK)gas back Kit and observe after 24-48 hrs in anaerobic conditions.

Gram stain reagent and Malachite Green: For morphological study.

Identification was performed by the fermentation of glucose, lactose, maltose and sucrose, in peptone yeast extract (PY) broth, and indole production; gelatin, esculin and starch hydrolysis ,and motility[15] .To further confirmation utilized API 20A kits (bio Mérieux).

API 20A KIT(bio Merieux ,Inc.USA ). System for the identification of anaerobes. The API 20 A system enables 21 tests to be carried out quickly and easily for the biochemical identification of anaerobes(table5). Other tests such as colonial and microscopic morphology, Gram stain, Malachite Green. Should be performed and the results used to confirm or complete the identification.[16].

Statistical Analysis

The Statistical Analysis System- SAS [17] was used to effect of different factors in study parameters. Chi-square test was used to significant compare between percentage in this study.

**Results and Discussion:** 

#### **Isolation and Identification:**

After 48 hrs *C. difficile* colonies grow circular, raised, opaque grey-white, sometimes with irregular borders, and(2-4)mm in diameter. In addition ,under UV light they produce a characteristic fluorescence. but with no signs of hemolysis. Well-grown cultures have a characteristic "horse-stable-like" odour. [18]. (fig:1).

Gram stain: positive bacillus cell shape, Arrangement: single or chains(2-6)cell sub terminal endo spore forming(fig2,3,4). API 20A strips used for identification of *C. difficile*. After 48 hrs of incubation all the isolates was confirmed.(table:4)and (fig5).



Figure(1). Typical C.difficile colonies on

CCFA containing blood.



Figure(2). Gram positive (1000x)



Figure(3):spore forming bacteria.1000x Figure(4):Malachite green stain spore.1000x



Figure(5):API 20A strip used for identification of *C. difficile*. After 48 hrs of incubation all the isolates was confirmed.

مجلة كلية التربية الأساسية - 60 - المجلد 21- العدد 87 - 2015

Api20A identification results of 51 isolate.						
	Test	results	No. positive	No. negative		
	IND		0	51		
	URE		0	51		
	GLU	+	51	0		
	MAN	±	5	1		
	LAC		0	51		
	SAC		0	51		
	MAL		0	51		
	SAL	ŧ	5	46		
	XYL		0	51		
	ARA		0	51		
	GEL	+	51	0		
	ESC	+	51	0		
	GLY		0	51		
	CEL	ŧ	2	49		
	MNE	ŧ	50	1		
	MLZ	+	51	0		
	RAF		0	51		
	SOR	ŧ	1	50		
	RHA	ŧ	1	50		
	TRE	±	1	50		
	CAT		0	51		
	SPOR	+	51	0		
	GRAM	+	51	0		
	COCC		0	51		

 Table (4). Api20A identification results of 51 isolate.

*C. difficile* were isolated by 51 (21.25)% from a total of 240 stool samples collected from children having diarrhea. The isolation percent from age $\leq$ 5 years was18.75% while it in age >5was2.5%(table:1)and(fig:1), and it was found that this percent decreased with age, it formed higher isolation percentage in ages<5 years. As a compare with controle, the difference was non statistically significant (*P*< 0.05).

Table 1. Distribution of study sample according to age groups

Age groups	Patient		Control		Chi-
years	Sample No.	Positive No.	Sample No.	Positive No.	square- $\chi^2$
		(%)		(%)	
≥ 5	216	45 (18.75)	60	12 (15.00)	0.849 NS
6-10	15	6 (2.50)	10	0 (0.00)	0.851 NS
11-15	9	0 (0.00)	10	0 (0.00)	0.00 NS
Total	240	51 (21.25)	80	12 (15.00)	2.077 NS
Chi-square-		6.71 **		5.724 *	
$\chi^2$					
* (P<0.05), ** (P<0.01), NS: Non-significant.					

- 61 - المبلد 21- العدد 87 - 61 -

مجلة كلية التربية الأساسية



Figure 1. Distribution of sample study according to age group (%)

The isolation percent from age <1year was 15%(table:2)and(fig:2), while in age>1year was 6.25 %.(table:1, 2). As a compare with controle, the difference was non statistically significant (P< 0.05).

Age group	Patient		Control		Chi-
years	Sample No.	Positive No. (%)	Sample No.	Positive No. (%)	square- $\chi^2$
> 1	147	36 (15.00)	20	11 (13.75)	0.637 NS
1-2	48	6 (2.50)	20	1 (1.25)	0.422 NS
2-3	9	2 (0.83)	10	0 (0.00)	0.048 NS
3-4	6	1 (0.42)	5	0 (0.00)	0.009 NS
4-5	6	0 (0.00)	5	0 (0.00)	0.00 NS
Total	216	45 (18.75)	60	12 (15.00)	0.804 NS
Chi-square- $\chi^2$		6.057 **		4.739 *	
* (P<0.05), ** (P<0.01), NS: Non-significant.					

Table 2. Distribution of study samples according to age groups<5 years.

ة الأساسية - 62 - المجاد 21- العدد 87- 2015



Figure 2. Distribution of sample study according to age group (%)

It was found that this percent decreased with age get older. This study shows that females were more infected than males (table:3)and(fig:3).

Table 3. Distribution	n of sample s	study according to sex
-----------------------	---------------	------------------------

Age (group)	Patient		Control		Chi-square- $\chi^2$
	Sample No.	Positive No.	Sample No.	Positive No.	
	(%)	(%)	(%)	(%)	
Male	102 (42.50)	21 (8.75)	36 (45.00)	5 (6.25)	0.144 NS
Female	138 (57.20)	30 (12.60)	44 (55.00)	7 (8.75)	0.208 NS
Total	240 (100)	51 (21.25)	80 (100)	12 (15.00)	0.963 NS
Chi-square- $\chi^2$	5.612 *	1.82 NS	4.853 *	1.579 NS	
* (P<0.05), NS: Non-significant.					



Figure 3. Distribution of sample study according to sex (%)

- 63 - المبلد 21 - 11هـ - 63

مجلة كلية التربية الأساسية

This result agrees with previous studies in Iraq [19] who isolated 13%, in age<1 year10% and in age>1 year 3% of *C. difficile* from children suffering from diarrhea on medication with antibiotic, while in healthy children 12%. And with another study who isolation percent was 17 % under 2 years[20].

Our study disagreement with another study was isolated 5.5% of children under 5 years and no healthy children harbored this microorganisms[21] .And disagreement with [22] was isolated 3% <15 years, also females were infected more than males [22] . This is an indicator that CDI has become an increasingly common infection and has shown an increase in severity over the past several years. Our goal with this study is that physicians will be more aware of the seriousness of AAD and be vigilant about quick diagnosis and appropriate treatment.

*C. difficile* is the cause of approximately(25–30)% of all cases of (AAD) [23]. The growth of infection may be caused by multiple factors including inappropriate antibiotic usage, poor standards of environmental cleanliness, changes in infection control practices, large outbreaks of *C. difficile* infection in hospitals, alteration of circulating strains of *C. difficile*, and spread of hyper virulent strains. [23].

Detection of high-risk populations could be helpful for prompt diagnosis and consequent treatment of patients suffering from *C*. *difficile* infection. Current treatments for *C*. *difficile* infection consist of supportive care, discontinuing the unnecessary antibiotic, and specific antimicrobial therapy.[24].

The purpose of this report is to evaluate the performance of the API 20A anaerobe system in definitively identifying strains of *C. difficile*., and to further confirmation utilized API 20A kits (bioMérieux).[25].

## References

- 1-Marie Céline Zanella Terrier, Martine Louis Simonet, Philippe Bichard, Jean Louis Frossard.(2014). Recurrent *Clostridium difficile* infections: The importance of the intestinal microbiota. World J Gastroenterol 2014 June 21; 20(23): 7416-7423.
- 2- Mattila E, Seppala R, et al. (2012). Fecal Transplantation, Through Colonoscopy, Is Effective Therapy for Recurrent *Clostridium difficile* Infection., GASTROENTEROLOGY..142:490–496.
- 3- Sahil Khanna and Darrell S. Pardi.(2014). *Clostridium difficile* infection: management strategies for a difficult disease. Ther Adv Gastroenterol 2014, Vol. 7(2) 72–86.

- 4- Shih-Ta Shang, Sheng-Kang Chiu, Ning-Chi Wang, and Jung-Chung Lin.(2013). *Clostridium difficile*–associated Diarrhea: Brief Review and Update of Medical Management. J Intern Med Taiwan 2013; 24: 309-316).
- 5- Victor R. C. Merino, Viviane Nakano, Sydney M. Finegold, andMario J. Avila-Campos.(2014). Genes Encoding Toxin of *Clostridium difficile* in Children with and without Diarrhea. Hindawi Publishing Corporation Scientifica Volume 2014, Article ID 594014, 4 pages.
- 6- Rousseau, C., Poilane, I., Pontual, L.D., Maherault, A., Le Monnier, A., Collignon, A. (2012) Clostridium difficile carriage in healthy infants in the community: A potential reservoir for pathogenic strains. Clin. Infect. Dis. 10: 1093.
- 7- Saad NM, Amin WF and Shaker EM (2013) Detection of toxigenic *Clostridium difficile* in powdered infant and follow-up formulae in Egypt, Veterinary World 6(11): 862-864.
- 8-Khanna, S. and Pardi, D. (2012). *Clostridium difficile* infection: new insights into management. Mayo Clin Proc 87: 1106–1117.
- 9-Jöbst, M., Heuberger, S., Indra, A., Nepf, R., Köfer, J., Wagner, M. (2010) *Clostridium difficile* in raw products of animal origin. Int. J. Food Microbiol. 138: 172-175.
- 10- Hannah GL. and Brandi L., (2010). Clostridium difficile in Food and Domestic Animals: A New Foodborne Pathogen? Clinical Infectious Diseases 51(5):577–582
- Gordon E. Schutze, MD, Rodney E. Willoughby, MD.(1013). *Clostridium difficile* Infection in Infants and Children. American Academy of Pediatrics (2013). *Pediatrics* Vol. 131 No. 1 January 1, 2013 pp. 196 -200.
- 12-Awanes ,A.S.(1997).Isolation and Identification of *Clostridium difficile* from children in Baghdad .MSC thesis, college of science ,university of Baghdad.
- 13-Luis Alcala,(2013). Laboratory tests for diagnosis of *Clostridium difficile* infection: Past, present, and future. Enferm Infecc Microbiol Clin. 2013;31(2):65–67).
- 14-Bo-Moon Shin, M.D. and Eun Joo Lee, M.T.(2014). Comparison of ChromID Agar and *Clostridium difficile* Selective Agar for Effective Isolation of *C. difficile* from Stool Specimens. Ann Lab Med 2014;34:15-19.
- 15-Holdeman LV, Moore WEC( 1977). Anaerobe Laboratory Manual, 4th ed., Anaerobe Laboratory, Virginia Polytechnic Polytechnic Institute and State University, Blacksburg, 200 pp.
- 16- Versalovic J., Caroll K.C., Funke G., Jorgensen J.H., Landry M.L., Warnock D.W(2011). Manual of clinical Microbiology. 10th Edition . American Society for Microbiology, Washington, D.C.
- 17- SAS. 2012. Statistical Analysis System, User's Guide. Statistical. Version 9.1<sup>th</sup> ed. SAS. Inst. Inc. Cary. N.C. USA.

- 65 ـ المجاد 21- العدد 87 - 2015

- 18- Kerin L. T, Diane M. C, Eliza S. L, Vreni C .M and Ellie J. C. (2013) .Goldstein. Evaluation of Cycloserine-Cefoxitin Fructose Agar (CCFA), CCFA with Horse Blood and Taurocholate, and Cycloserine-Cefoxitin Mannitol Broth with Taurocholate and Lysozyme for Recovery of *Clostridium difficile* Isolates from Fecal Samples J. Clin. Microbiol. 2013, 51(9):3094.
- 19- Ghanim H.Majeed,(2003).Isolation ,Identification and Toxin production From *Clostridium difficile* In patients with diarrhea and colitis.p:121.ph.D Thesis, College of Science ,Baghdad University.
- 20- Amera M. Al-Rawi,(2005). Isolation and Identification of *Clostridium difficile* from Patients with Colitis and Diarrhae in Ninevah Governorate. AL-Rafedaen science JOR.V16,N8,P:32-41.
- 21- Claudia EA F, Viviane N ,Edison L D,Mario J A. (2003) Prevalence of *Clostridium* spp. and *Clostridium difficile* in Children with Acute Diarrhea in São Paulo City, Brazil. Mem Inst Oswaldo Cruz, Rio de Janeiro, Vol. 98(4): 451-454, June 2003.
- 22-Diea A. Younis,(2008). Epidemiology of hospitilized patients in *Clostridium difficile* Mosul city.AL-TAQANI Journal,Volume21,page:A44-A48.
- 23- Cohen SH, Gerding DN, Johnson S, (2010).Clinical practice guidelines for *Clostridium difficile* infection in adults: 2010 update by the society for healthcare epidemiology of America SHEA) and the infectious diseases society of America(IDSA). Infect Control Hosp Epidemiol 2010; 31: 431-55.
- 24-Mehdi Goudarzi, Sima Sadat Seyedjavadi, Hossein Goudarzi, Elnaz Mehdizadeh Aghdam, and Saeed Nazeri.(2014). *Clostridium difficile* Infection: Epidemiology, Pathogenesis, Risk Factors, and Therapeutic Options. Hindawi Publishing Corporation Scientifica Volume 2014, Article ID 916826, 9 pages.
- 25- MARY E. G., CAROL J. S., DALE N. G, CONNIE R. G, AND LANCE R. P ('1984). Evaluation of the 24-h API 20A Anaerobe System for Identification of *Clostridium dijficile*. JOURNAL OF CLINICAL MICROBIOLOGY, June 1984, p. 915-916.

مبلة كلية التربية الأساسية - 66 - المبلد 21- العدد 87- 2015

Table:5 READING TABLE API20A						
TESTS	ACTIVE INGREDIENTS	REACTIONS/ENZYMES	RESULTS			
	II (OREDIEI (II)		NEGATIVE	POSITIVE		
IND	L-tryptophane	INDole formation	<u>XYL - mix / 2-3 min</u>	+EHR / 5 min		
UDE		LIDEasa	<u>Yellow</u>	<u>Red</u>		
OKE			yenow-orange	Ieu DCD		
GLU	D-glucose	acidification (GLUcose)	<u>BCb</u>	<u>BCP</u>		
MAN	D-Indimitor D-lactose	acidification (MANIII0)				
LAC	(bovine origin)	aciumeation (LACtose)				
SAC	D-saccharose (sucrose)	· · · 1' C' · · · · ·	purple	Yellow/yellow-		
MAL	D-maltose	acidification (SACabarasa)	1 1	green		
SAL	salicin	(SACCHAROSE)		0		
XYL	D-xylose	acidification (SAL icin)				
ARA	L-arabinose	acidification (XYLose)				
		acidification				
		(ARAbinose)				
GEL	gelatin	hydrolysis (protease)	no diffusion of	diffusion		
	(bovine origin)	(GELatin)	pigment (1)	of black pigment (1)		
ESC	esculin	hydrolysis (ß-glucosidase)	yellow (2)	brown-black (2)		
	ferric citrate	(ESCulin)	in UV (365	nm		
			nm	no fluorescence		
CLV	alvaaral	agidification (CI Vegral)	nuorescence	DCD		
CEL	D-cellobiose	acidification (OL I CEIOI)	ber	<u>bcr</u>		
MNE	D-mannose	(CEL lobiose)				
MLZ	D-melezitose	acidification (ManNosE)				
RAF	D-raffinose	acidification	purpule	Yellow/yellow-		
SOR	D-sorbitol	(MeLeZitose)		green		
RHA	L-rhamnose	acidification				
TRE	D-trehalose	(RAFfinose)				
		acidification (SORbitol)				
		acidification				
		(RHAmnose)				
САТ		CATalase	After 30 min	H2O2 in a positive		
CAI		CATalase	in air	<u>tube</u>		
			No bubble	bubble		
				<u></u>		
SPOR		spores	absent	present		
GRAM		Gram reaction	pink	violet		
COCC		morphology	rod	coccus		

- 67 - المجلد 21- العدد 87 - 67 -

مجلة كلية التربية الأساسية

Isolation and Identification of Clostridium difficile from Antibiotic-Associated Diarrhea and Colitis in Iraqi children, using API20A Anaerobic system..... Mona Turkey AL-Mossawei, Luma Yousif Mahdi عزل وتشخيص جرثومة Clostridium difficile من أطفال عزل وتشخيص يعانون الإسبهال والتهاب القولون والمصاحب للمضادات API20A anaerobic system

لمى يوسف مهدي<sup>2</sup>

منى تركي الموسوي<sup>1</sup>

جامعة بغداد - كلية العلوم للبنات / قسم علوم الحياة.

الخلاصة:

تضمنت الدراسة عزل وتشخيص Clostridium difficile من الأطفال المصابين بحالات الإسهال والتهاب القولون الناجمين عن تناول المضادات الحيوية وبأعمار من بعد الولادة ولغاية 15 سنة.تم جمع العينات من الفترة مابين الاول من حزيران 2013 ولغاية الاول من نيسان 2014 .اضافة الى جمع 80 عينة من اطفال اصحاء كمجموعة سيطرة.اعتمدت الاختبارات المظهرية واستخدام نظام API20A في التشخيص وتعزيز الاختبارات الكيمياحيوية. بينت النتائج أن نسبة عزلها من عينات الإسهال عند الاطفال البالغة 240 عينة كان (21.25%).اوضحت الدراسة ان نسبة اصابة الاناث اعلى بقليل من الذكور وعدم ظهور فروق معنوية بين المرضى والسيطرة.وعند دراسة تأثير العمر على نسبة عزلها تبين ان النسبة تقل مع زيادة العمر , إذ بلغت أعلى نسبة عزل(18.75%) عند الفئة العمرية الاقل من 5سنوات.